Microbiology Research Journal International



30(9): 70-77, 2020; Article no.MRJI.62951 ISSN: 2456-7043 (Past name: British Microbiology Research Journal, Past ISSN: 2231-0886, NLM ID: 101608140)

Bacteriological Profile of Nosocomial Infections in Visceral Surgery at the CNHU-HKM of Cotonou in Republic of Benin

Aïkou Nicolas^{1*}, A. Ahoyo Theodora², Degbe Sah Cyriaque³, Coulibaly Founzégué Amadou⁴, Sezan Alphonse⁵, Edorh A. Patrick⁵ and R. A. Lapo⁶

¹Department of Human Biology, National University of Sciences, Engineering and Mathematics, Laboratory of Clinical Biochemistry and Medical Microbiology, Benin.
²Abomey Calavi University, General Medical Microbiology and Hospital Hygiene Abomey Calavi Politechnical School, Human Biology Engineering, Benin.
³Regional Institute of Public Health Ouidah, Benin.
⁴Pasteur Institute Abidjan, Cote-d'ivoire.
⁵FAST/UAC, Benin.
⁶Inter-state School of Science and Veterinary Medicine, Dakar, Senegal.

Authors' contributions

This work was carried out in collaboration among all the authors. Authors AN, AAT and DSC designed the study. Author AN wrote the manuscript. Authors CFA, SA, EAP and RAL managed the statistical analyzes of the study. All authors have read and approved the final manuscript.

Article Information

DOI: 10.9734/MRJI/2020/v30i930265 <u>Editor(s):</u> (1) Dr. Ana Cláudia Coelho, University of Trás-os-Montes and Alto Douro, Portugal. <u>Reviewers:</u> (1) Kishore Chandra Thakur, India. (2) Silvia Farouk Shalaby, Cairo University, Egypt. Complete Peer review History: <u>http://www.sdiarticle4.com/review-history/62951</u>

Original Research Article

Received 15 September 2020 Accepted 24 November 2020 Published 11 December 2020

ABSTRACT

Wound suppuration is the formation and accumulation of pus in the soft tissue of the wound. As a rule, the natural inflammatory reaction disappears within three to five days and later the wound heals without any character. During this time, in some cases the inflammatory reaction in the wound exceeds the limits of the physiological norm due to the external supply of bacterial germs,

*Corresponding author: E-mail: aikounicolas@yahoo.fr, nicolashoundjo@gmail.com;

Nicolas et al.; MRJI, 30(9): 70-77, 2020; Article no.MRJI.62951

which leads to the formation of pus, exaggeratedly prolonging the healing process and in turn leading to serious consequences. This study aims to identify the bacteria involved in these suppurations, to study their antibiotic resistance profiles and to review the role of the nurses taking care of these patients. The study was conducted at the Menontin zone hospital from May 15 to June 21, 2013. Fifty-one patients of all ages and sexes with surgical and traumatic discharge wounds were sampled and 30 hand samples were taken from the nurses' hands before and after dressing. From the results obtained, 76 bacteria were identified, including 35 cocci and 41 bacilli. 29% of the bacteria found were Pseudomonas aeruginosa, 28% Staphylococcus aureus, 14% Escherichia coli and others. Hand samples of 10 bacteria were isolated including 4 Pseudomonas aeruginosa, 3Staphylococcus DNase-negative 2 Klebsiella spp and 1 Staphylococcus aureus. The strains isolated were particularly resistant to the antibiotics tested more specifically to β-lactam, Gentamycin Erythromycin and Colistin. The majority of strains have good sensitivity to ciprofloxacin and Fosfomycin. In view of these results, it is necessary to draw the attention of patients and nurses to the reality of the existence of these bacteria, the adequate intake of antibiotics preferably after an antibiotic test, hand washing and the use of appropriate sterile materials before and after any dressing.

Keywords: Nosocomials; bacterial strains; bacteriological profile; antibiotic; sensitivity; resistance.

1. INTRODUCTION

Bacteria are microscopic microorganisms that populate our environment, providing for some beneficial actions and for others harmful effects on the human organism depending on their location and pathogenicity [1]. In hospitals or clinics there are many sources of infectious germs responsible for diseases whose frequency varies according to the nature and structure of the hospital on the one hand and the activity of the hospital units and the quality of the health care team on the other [2] without forgetting the quality of the hygiene of the operating rooms and the medical equipment used [3]. The irrational use of antibiotics by patients themselves, combined with the lack of respect for hygiene rules, are all factors in the resurgence of these hospital infections. The problems they cause are in terms of morbidity, mortality and additional costs [4]. It is estimated that 60% of hospital infections worldwide are due to multi-resistant bacteria [5], which calls for more stringent hospital infection control measures. In Benin, for example, during our final training courses at the Menontin hospital laboratory, pus analyses carried out showed that the antibiotics prescribed by the clinician as a preventive measure, without an antibiotic susceptibility test, are mostly ineffective in preventing suppurative infections of open wounds related to traumatic accidents or surgical procedures. We have noticed that the dressing department of this hospital is overwhelmed by the management of traffic accidents, caesarean sections and other wounds, which have greatly contributed to the

increase in the number of dressings to be performed by the nurses. Are they still taking precautions to limit the transfer of germs, when hygiene and infection control must be carefully monitored?

2. STUDY FRAMEWORK AND METHOD

2.1 Study Framework

For this study, we chose the Menontin hospital located in the 9th district of the urban commune of Cotonou, in the Ménontin lot 2130A neighborhood, not far from RNIE 2.

2.2 Study Method

51 pus samples were collected from any patient admitted to Menontin Hospital of any age and sex after their consent was obtained. These patients had a skin wound (superficial or deep) with suppurating pus, with two sterile swabs, one of which will be used for direct examination (fresh) and the other put in sterile peptone water and used to make the culture.

30 hand samples were taken from the nurses' hands before and after they had applied three dressings. The sample was collected with a sterile swab (moistened with sterile peptone water) applied to the palms of the hands and interdigital spaces.

The analysis of one sample took 72 hours over 3 days:

1st day: Fresh state, gram staining and culture according to the germ found on the gram slide. According to the present germ, were sown:

- For gram-positive cocci: chapman agar, fresh sheep blood agar 5% and D-cocosal agar.
- For gram negative bacilli: EMB agar, Schaedler agar enriched with fresh sheep blood 5%.
- The boxes were incubated at 37°C in the oven for 24 hours, those with blood were incubated anaerobically at 37°C for 24 hours.

2nd day: If there is a culture, whatever the colonies, a gram control has been done as well as the following tests:

- For gram-positive cocci: search for catalase and if positive, search for respiratory type and then search for DNase.
- For gram-negative bacilli: search for oxidase and if the latter is positive a seeding of the colony is done on MH agar and on Basal Medium plus glucose for the identification of pseudomonas. For gramnegative bacilli with negative oxidase, a mini biochemical gallery was inoculated on Urea-indole, Kligler, Manitol-Mobility, Simmons Citrate and then on VP-RM medium.

Still for this J2, the antibiogram was carried out with :

- Gram-positive cocci, the following TBA discs: Ampicilin, Amoxicillin + Clavulanic acid, Carbenicillin, Oxacillin, Cefotaxime, Ceftriaxone, Gentamycin, Erytromycin, Nitroxoline, Ciprofloxacin, Sulfadoxin, and Fosfomycin.
- Gram-negative bacilli: Beta-Lactamines (Ampicillin, Amoxicillin + Clavulanic acid, lipenem, Aztreonam, Cefalotin, Cefotaxime, Ceftriaxone); Aminosides (Gentamycin); Quinolones (Ciprofloxacin); Sulfonamides (Sulfadoxin); Polypeptides (Colistin); and Fosfomycin.

All media were incubated at 37° in an oven for 24 hours.

3rd day:_reading of inoculated media from J2 with finalization of identification tests including DNase, Indole, TDA; VP and MR as well as TBA

reading by measuring antibiotic inhibition diameters and comparing them to the provided reading scale.

3. RESULTS AND DISCUSSION

3.1 Result

3.1.1 Samples taken with positivity cases

There are 51 suppurative wounds collected from patients and 30 are hand samples taken, 22 of which were taken before dressing and 18 after dressing. Fig. 1 shows the cases of positivity of these samples.

3.1.2 Survey results

From the individual patient information sheets we noticed that more than half of them (29/51) were already under antibiotic treatment (Beta-Lactamines, Aminosides and Quinolones) and even with imidazoles. Combinations of treatment are sometimes performed.

3.1.3 Isolated germs and their prevalence

The germs isolated in wound healing during our study totalled 76, of which 35 (46%) were grampositive cocci and 41 (54%) gram-negative bacilli.

For the hand samples 10 germs were isolated, 6 before dressing and 4 after dressing. Their identities and numbers are shown in the Table 1.

3.1.4 The resistance profile of these germs

Regarding the resistance profile of the bacteria isolated from the bacterial species showed strong resistance to most of the antibiotics tested apart from Fosfomycin which is active on almost all strains.

3.2 Discussion

The purpose of this study is first to identify the bacteria involved in wound healing, to establish the resistance profile of these bacteria and then to examine more closely the likely role of nurses in this fact. We had 51 suppurative wound samples, 50 of which were infected with bacteria, to support the fact that pus is generally the result of an inflammatory process due to an interaction between tissue and pyogenic germs [6]. From the nurses' hand samples, cases positive for

germs have been recorded, including 6 before dressing and 4 after, which may be linked to poor hand hygiene, in line with Cooper's statement that "the human body is not sterile", and aseptic hand washing could reduce or even eliminate residential flora for a period of time [7,8].

76 bacteria were isolated from suppurative wounds with a predominance of gram-negative bacilli (54%) versus 46% of gram-positive Cocci, a result similar to that obtained by Rao et al. [9]. Of the bacteria identified in our study, *Pseudomonas aeruginosa was* the most common of the bacilli (29%) and *Staphylococcus aureus of the* cocci (28%), which is similar to the study by Slekovec et al. [10] and Hani et al. [11]. For these authors, these two bacteria are the most isolated from suppurative wounds, with Mohammed et al. showing a good inclination towards *Staphylococcus aureus* [12].

From the hands of the nurses we isolated Pseudomonas, Staphylococci and Enterobacteriaceae before and after the dressing. This shows that nurses are likely to contaminate patients' wounds, as Khoury says. The same author adds that the work environment can also be a source of infection [13]. Jeurissen et al. have shown that the quality of the bandage may promote the penetration of germs, leading to wound healing [14].

As for the antibiotic resistance profiles of these germs, we will start with Pseudomonas. The strains isolated were highly resistant to β-lactam antibiotics but highly susceptible to Ciprofloxacin, Ceftriaxone and Fosfomycin. These results are similar to those obtained by Hani and AL [11]. Mohammed et al. [12], as in our study, found that Pseudomonas isolated from wounds were sensitive to Ciproflocacin. Although it is reported that Pseudomonas are frequently isolated in wound infections and are highly resistant [15]; the emergence of these resistant strains, especially to antibiotics of last resort such as Colistin and Imipenem, should be of concern. Indeed, all strains isolated have been resistant to these antibiotics, but Auajjar et al. and Mesaros et al. have shown that Pseudomonas are highly to these antibiotics susceptible [15,16]. According to Rao et al. and Etok et al. [9,17], Imipenem is the most active. But according to Randrianirina et al. less than 50% of the strains would be sensitive to it [18]. Pseudomonas are bacteria that accumulate numerous mechanisms of antibiotic resistance. This would explain our results regarding the high resistance of the strains to Imipenem (13/22) and Colistin (22/22).

Next comes Staphylococcus aureus, which is generally the most common cause of surgical and traumatic wound infections [19]. The emergence of multi-resistant strains is thought to be one of the main causes of delayed and difficult healing of suppurative wounds. In our study, all strains of staphylococci isolated are resistant to Oxacillin (1µg) and there is a high resistance to sulfonamides. Our results are similar to those obtained by Sani et al. [19], Etok et al. [20] and Ogounshe et al. [21] in terms of the proportions of MRSA. Sani et al. have also shown in studies in Niger that staphylococci are highly resistant to sulfonamides. In contrast, Anas [22] obtained 50% methi-R pathogenic Staphylococci and 25% sulfadoxin-resistant Staphylococci. Sina et al. [23] in Benin isolated pus, only 42.85% MRSA and 51.42% of the sulfadoxin-resistant strains. The high resistance observed to Sulfonamides, Macrolides and Aminosides in streptococci, the second most common cocci involved in wound infections after staphylococci [24], was also demonstrated by Lopardo and Sood et al. [24,25] but in our study they showed a high sensitivity to Ampicillin. The high sensitivity of cocci to Fosfomycin is not surprising given that many studies in recent years have shown that this antibiotic is effective on more than 80% of cocci [26,27,28,29].

Enterobacteriaceae make up a quarter (25%) of the bacteria isolated. The high resistance extended to almost all families of antibiotics is similar to that observed by Randrianirina et al and Sani et al. [18,19]. The majority of the Enterobacteriaceae isolated from our samples are sensitive to Ciprofloxacin, Sani et al; Tumane and Wasnik [18,30] obtained similar results while Mohammed et al obtained high resistance to this antibiotic.

We noticed that in vitro, the antibiotics prescribed to patients were virtually inactive on the strains taken in isolation. This resistance of the different strains studied could be related to the fragile state of the patients sampled or even due to wound infection by nosocomial strains [31] accidentally colonizing the wounds through work equipment, gowns, bed sheets, the nurse's hands, to name but a few reasons. In addition, it could be natural [31] for some strains or acquired [31,32,33] for others due to the duration of treatment or inappropriate antibiotic prescriptions [34]. Hence the need for judicious use of antibiotics to reduce hospital-acquired infections in general and in the management of wound infections in particular [35].

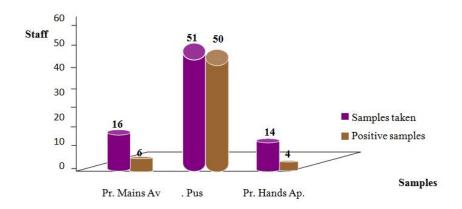


Fig. 1. Number of positive samples as a function of samples taken

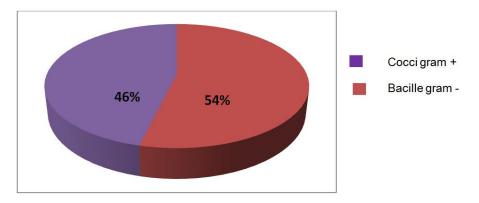
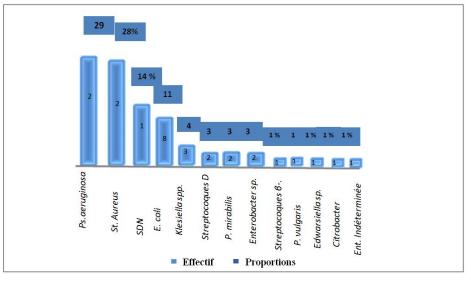


Fig. 2. Distribution of isolated germs as a function of Gram and Fig. 3 reveals the identity of its germs and their prevalences



Legend: SDN: Staphylococcus with DNase Negative - Ent: Enterobacterium

Fig. 3. Proportions of Bacteria Isolated from Suppurative Wounds

		Hand samples		
Isolated bacteria		Before dressing	After dressing	Total
No Pathogen	SDN	02	01	03
Pathogen	St. Aureus	01	00	01
-	K. pneumoneae	01	01	02
	Ps. Aeruginosa	02	02	04
Total		06	04	10

Table 1. Hand samples

4. CONCLUSION

At the end of our study and in view of the results obtained we can say that the following bacteria : Pseudomonas aeruginosa, Staphylococcus DNase-negative Staphylococci, aureus. Escherichia coli, Klebsiella (K.oxytoca, K. pneumoniae). Streptococci. Proteus. Enterobacter, and Citrobacter are those involved in the suppuration of skin wounds and most of these germs were found on the hands of the nurses who dress these wounds. This underlines the need to promote hygiene measures, with aseptic hand washing being at the forefront of these measures, without overlooking the fact that the dressing kit is single-use. Isolated bacteria have shown strong resistance to several tested and commonly prescribed antibiotics including β lactam antibiotics, aminoglycosides, macrolides and polypeptides. Only fosfomycin and guinolone antibiotics were active on the majority of the strains isolated. It is therefore necessary to develop a program to fight against nosocomial infections and against multiple bacterial resistances involving the entire health care chain: Ministry of Health, clinicians, nurses, biologists; and pharmacists. The promotion of our natural essences would be, above all, a great help in the fight against multi-resistant germs.

CONSENT

51 pus samples were collected from any patient admitted to Menontin Hospital of any age and sex after their consent was obtained.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

 Naïrouz B. Etude de l'activité antibactérienne des huiles infusées de quatre plantes médicinales connaisseurs comme aliments. Thesis presented for the magister diploma in pharmacochemistry, Mentouri University of Constantine Institute of Chemistry. 2005;5.

- Association of Professors of Infectious and Tropical Pathologies (APIT). (). Nosocomial infections. Infectious Diseases. 12th Edition. 1992;311-316.
- WWW pour la science.fr> infection medicine in hospital.
- Awari A, Nighute S, Deorukhkar S. Surgical wound infections: A prospective Hospital based study. Journal of Clinical and Diagnostic Research. (Suppl-2), 2011;5(7): 1367-1370.
- Carle S. Antibiotic resistance: an important public health issue! Pharmactuel. 2009; 42(2):6- 21.
- Da Costa J. C. Suppuration and Abscess. Modern Surgery, Chapter 6. 4th Edition. 1903;53. Available:http://jde.jefferson.edu/dacosta_mo

dernsurgery/537. WHO. Hand Hygiene: Technical Reference Manual. 2010;8.

- Center for coordination of the fight against nosocomial infections of the Paris-North inter-region. Hand hygiene: Guide to good practice". ^{3rd} Edition. 71 pages; 2001.
- Rao R, Sumathi S, Anuradha K, Venkatesh D, Krishna S. Bactériology of postoperative wound infections. International Journal of Pharmaceutical and Biomedical Research. 2013;4(2):72-76.
- Slekovec C, Faivre B, Humbert P., Bertrand X, Hocquet D, Pazart L, Talon D.. Les soins des plaies chroniques entraînent une contamination bactérienne de l'environnement. Annales de dermatologie et de vénéréologie. 2012;139:798–802.
- 11. Masaadeh HA, Jaran AS. Incident of Pseudomonas aeruginosa in post-operative wound infection. American Journal of Infectious Diseases. 2009;5(1):1-6.
- Mohammed A, Adeshina GO, Ibrahim YK. Incidence and antibiotic susceptibility pattern of bacterial isolates from wound infections in

a tertiary hospital. Journal of Pharmaceutical Research. 2013;12(4): 617-621.

- Khoury L. L'indeminisation des victimes d'une infection nosocomiale au Québec: les leçons du droit français. Les Cahiers de droit. 2004;45(4):619-657
- Jeurissen A, Hendrick R, Beesemans K, Van Thielen A, Cossey V, Schuermans A. Safety of non-woven polypropylene surgical adhesive drapes to prevent wound infection. BMC Proceedings from International Conference on Prevention ET Infection Control, Geneva. 20115(6): 190.
- Mesaros N, Nordmann P, Plésiat P, Roussel-Delvallez M, VanEldere J, Glupczynski Y, VanLaethem Y, Jocobs F, Lebecque P, Malfroot A, Tulkens PM, Van bambeke F. Pseudomonas aeruginosa: Resistance and therapeutic options at the dawn of the second millennium. Louvain Medical. 2007;126 (8):305-316.
- Auajjar Nr, Attarassi B, Elhaloui NE, Badoc A. Multiresistance aux antibiotiques de Pseudomonas aeruginosa, P. fluorescens et *Staphylococcus* aureus et survie sur divers tissus hospitaliers. Bull. Soc. Pharm. Bordeaux. 2006;145:61-76.
- Etok CA, Edem EN, Ochang E. Aetiology and antimicrobial studies of surgical wound infections in University of Uyo Teaching Hospital (UUTH) Uyo, Akwalbom State, Nigeria. 2012;1:341.
 DOI:doi.10.4172/scientificreports.341
- Randrianirina F, Vaillant L, Ramarokoto CE, Rakotoarijaona A, Andriamanarivo ML, Razafimahandry HC, Randrianomenjanahary J, Raveloson JR, Hariniaina ER, Carod J-F, Talarmin A, Richard V. Antimicrobial resistance in pathogens causing nosocomial infections in surgery and intensive care wards in Antananarivo, Madagascar. J. Infect. Dev. Ctries. 2010;4(2):074-082.
- Sani RA, Garba SA, Oyewole OA. Antibiotic resistance profile of Gram negative bacteria isolated from surgical wounds in Minna, Bida, Kontagora and Sulejaareas of Niger State from surgical wounds ins Minna, Bida, Kontagora and Sulejaareas of Niger State ", American journal of Medicine and Medical Sciences. 2012;2(1):20-24.
- Etok CA, Edem EN, Ochang E. Aetiology and antimicrobial studies of surgical wound infections in University of Uyo Teaching Hospital (UUTH) Uyo, Akwalbom State, Nigeria. 2012;1:341.
 DOI:doi.10,4172/scientificreports.341

- Ogunshe AAO, Niemogha MT, Azum GN. ,Odikagbue AN. Microbiological evaluation of antibiotic resistance in bacterial flora from skin wounds. Journal of Pharmaceutical and Biomedical Sciences. 2012;22(06):1-7.
- 22. Anas M. Les infections nosocomiales à propos des 55 cas colliges au maroc", Thesis presented and publicly defended to obtain the degree of doctor of pharmacy, Université CheikH Anta Diop de Daka. 2002;3:5-11.
- Sina H, Baba-Moussa F, Ahoyo TA, Mousse W, Anagonou S, Gbenou JD, Prévost G, Kotchoni SO, Baba-Moussa L. Antibiotic susceptibility and toxinsproduction of *Staphylococcus* aureus isolated from clinical sambles from Benin. African Journal of Microbiology Research. 2011;5(18): 2797-2803.
- 24. Sood S, Malhotra M, Das BK, Kapil A. Enterococcal infections et anti microbial resistance. Indian Journal Medical Research. 2008 ;128 :111–121.
- Lopardo H. Antimicrobial resistance in βhemolytic streptococci in Argentina. Communicating current research and educational topics and trends in applied microbiology. A Méndeé-Vilas (ED.). 2007;794-798.
- Honderlick P, Cahen P, Gravisse J, Vignon D. What antibiotic sensitivity for bacteria causing urinary tract infections? What about fosfomycin and nitrofurans? DOI :doi.10.1016/j.patbio.2006.07 .016.
- 27. WHO/CDS/CSR/EPH. Prevention of nosocomial infections: guide to good practice", ^{2nd} edition. 2008;771.
- Kastoris AC, Rafailidis PI, Vouloumanou EK, Gkegkes ID, Falagas ME. Synergy of fosfomycin with other antibiotics for Grampositive and Gram-negative bacteria. Eur J Clin Pharmacol. 2008;66:359-368.
- 29. Karakasa, Turhanv. Problem of antibiotic resistance in patients with acute bacterialcystilis and treatment options. Turk J Med Sci; 2012;42(Sup.2):1542-15 43.
- Tumane PM, Wasnik DD. Occurrence of extended spectrum bêta - Lactamase producing Enterobacteriaceae causing wound infections. Asian Journal of Biomedical and Pharmaceutical Sciences. 2013;3(20):55-58.
- Dupont H, Plantefève G. Infections sévères à entérocoque en reanimation", Conférences d'actualisation, Editions scientifiques et médicales Elsevier SAS, and Sfar. 2002;541-554.

- 32. Frazee BW, Lynn J, Charlebois ED, Lambert, Lowery D, Perdreau-Remington F. High prevalence of methicillin-resistant *Staphylococcus* aureus in emergency department skin and soft tissue infection. Annalses Emergency Medical. 2005; 45(3):311-20
- 33. Wellington EMH, Boxall ABA., Cross P, Feil EJ, Gaze WH, Hawkey PM, Johnsonn-Rollings AS, Jones DK, Lee NM, Otten W, Thomas CM, Williams AP. The role of the natural environment in the emergence of Antibiotic resistance in Gram-negative

bacteria. Lancet Infect. Dis. 2013;13:155-65.

- Elhani D, Elhani I, Aouni M. Résistance chez les bacilles Gramnégatif : Où en sommesnous? La tunisie Medicale – 2012;90(10): 680-685.
- Singh V, Chauhan PK, Bodh UA, Kaushal K, Iqbal A. Isolation and antibiogram pattern of methicillin resistant *Staphylococcus* aureus causing wound infection. International. Journal of Analytical Pharmaceutical and Biomedical Sciences. 2012;1(1):18-21.

© 2020 Nicolas et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history: The peer review history for this paper can be accessed here: http://www.sdiarticle4.com/review-history/62951