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Phytochemical and Phytomineral Status of Spigelia anthelmia Linn Leaves

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Authors' contributions

This work was carried out in collaboration among all authors. Author OLA designed the work and conduct the laboratory analysis. Author POO performed the statistical analysis, wrote the protocol, and wrote the first draft of the manuscript. Author OSA managed some of the analyses of the study. Author EPC managed the literature searches. All authors read and approved the final manuscript.

Article Information

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Original Research Article

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ABSTRACT

Aims: *Spigelia anthelmia* is a medicinal plant used in the treatment of diverse diseases and contain plant-based natural bio-active constituents.

Study Design: Phytochemical and mineral analysis of *Spigelia anthelmia* leaves using standard analytical procedures.

Place and Duration of Study: Forestry Research Institute of Nigeria, between May 2019 and July 2019.

Methodology: Phytochemical and phytomineral status of *Spigelia anthelmia* leaves were investigated using standard analytical procedures. Phytochemicals screening/analysis examined include; alkaloids, flavonoids, saponin, tannin, phenolics, cardiac glycosides, phlobatannin and terpenoids. The atomic absorption spectrophotometer (AAS) was used to determine the minerals which are calcium (Ca), phosphorus (P), magnesium (Mg), iron (Fe), zinc (Zn), manganese (Mn), and copper (Cu), while flame photometer was used in determining potassium (K) and sodium (Na). Data were presented using descriptive statistics (mean and standard deviation).

Results: Result indicates the presence of phytochemicals in the leaves of *S. anthelmia*, the quantitative analysis shows: alkaloid (2.34mg/100g), flavonoid (6.13mg/100g), saponin

(18.12mg/100g), tannin (9.61mg/100g), phenol (4.61mg/100g), cardiac glycosides (1.89mg/100g), terpenoids (0.98mg/100g) and phlobatannin (0.32mg/100g). The macro elements reveals Na (0.022%), K (0.23%), Ca (0.48%), Mg (0.15%), P (0.035%), while the micro element shows that Fe (0.19%), Mn (0.003%), Cu (0.001%), Zn (0.004%).

Conclusion: The result suggests that *S. anthelmia* has a strong pharmacological prowess and could be used in the prevention and treatment of various health-related diseases.

Keywords: Bioactive; constituents; phytochemical; Spigelia anthelmia; oxidative.

1. INTRODUCTION

Herbal medicine is the use of herbs for their therapeutic or medicinal value [1]. The medicinal and pharmaceutical properties of medicinal plants are due to the type of chemical substance they produce and possess. These include primary metabolites that are utilized as food and give nutritional benefits and also secondary metabolites that exert physiological effects and give therapeutic benefits to plants. The term used for different chemical substances is called "constituents". The plant's constituents that have pharmacological properties are 'active constituents". The chemical constituents possessed by plants is very essential and widely studied because it helps in the production of most drugs used as medicine [2]. Secondary metabolites are the chemical components that are of medicinal benefits. The analysis of the plants chemical constituents can reveal the accumulation of specific compounds in some plants organ. The abundance and deficiency of such constituents largely depend on the specificity of plants, the extent of accumulation and the analytical method employed [3]. Phytochemicals are large chemical compounds naturally derived from plants, vegetables and fruits and are responsible for disease protection [4]. Based on their chemical structure, phytochemicals can fall into the following groups; flavonoids, saponin, alkaloids, tannin, steroids, phenols, terpenoids, cardiac glycosides [5]. Spigelia anthelmia is an annual weed that belongs to the genus of flowering plants in the Loganiaceae family. It is also known as pinkroot. It has a small purple flower with a feather lobed leaves [6]. The plant is mostly used for the expulsion of tapeworm and roundworm in the body. Also, it is used to treat people with chronic catarrh, difficulty in breathing, headache, throbbing pain, vermifuge and common cold [7]; Previous studies on different solvent extracts of S. anthelmia revealed the presence of varying presence of phytochemicals [8]. Studies on plantbased chemicals such as phytochemicals and minerals present in S. anthelmia, some of which

have been demonstrated to possess healthboosting properties. However little study has been done reporting the mineral status of *S*. *anthelmia*, and in view of this, this study aims at reporting the phytochemical status (both qualitative and quantitative) and the phytomineral composition of *S*. *anthelmia*.

2. METHODOLOGY

2.1 Collection and Preparation of Plant Materials

The leaf samples of *S. anthelmia* were collected within the premises of Forestry Research Institute of Nigeria, identified and authenticated by a taxonomist at the Forest Herbarium Ibadan and a voucher specimen (FHI 112462) was deposited. The leaves were air-dried at room temperature after which it was milled to powdered using a milling machine (Arthur H. Thomas Co. Phila, P.A. S.A). The fine powder was then packed into a tight container and kept for further use.





2.2 Phytochemical Evaluation

Qualitative screening and Quantitative phytochemical analysis were done to identify and quantify the chemical components present in the leaves of *S. anthelmia.* The bioactive constituents assayed includes; alkaloids,

saponin, flavonoids, tannin, phenolics, terpenoids, cardiac glycosides and phlobatannin. The phytochemical screening was evaluated on the powdered sample using simple standard procedures as described by Harborne [3], while the quantitatve phytochemical analysis was done according to the methods described by Mbaebie [9] using spectrophotometric and gravimetric principle.

2.3 Mineral Evaluation

1 g of the powdered sample was weighed in porcelain crucibles which were ignited in a muffle furnace at 550°C. The ash was dissolved in 3 ml of 3 M Hydrochloric (HCl) acid, transferred to 100 ml volumetric flask and diluted to the 100 ml mark. Potassium and sodium were analyzed using Flame Photometer while, iron, zinc, copper, manganese, and magnesium were analyzed using a Buck Scientific 210 atomic spectrophotometer, absorption phosphorus UV-Vis determined was using Spectrophotometer [10].

2.4 Data Analysis

Quantitative data were expressed as Mean ±SD of triplicate measurement; analysis of variance (ANOVA) was used to detect significant difference between mean measured of parameters of different species, while specific differences were identified using Least Significant Difference (LSD) statistical test at 5% level of probability. SPSS version 20 was used for the statistical analysis.

3. RESULTS

3.1 Qualitative Screening

The result presented in Table 1 shows the phytochemical screening of *S. anthelmia*. The result reveals the presence of a variety of secondary metabolites like saponin, tannin, flavonoid, terpenoids, phenols, alkaloids, phlobatannin and cardiac glycosides. The indication of these phytochemicals suggests that the plant *S. anthelmia* is a very potent plant that could be used in the treatment and prevention of various diseases.

3.2 Quantitative Analysis

The quantitative phytochemical analysis of S. anthelmia, as expressed in Table 2 reveals the levels of phytochemicals present in the leaves of S. anthelmia. The saponin content obtained in the study was the highest with a value of (18.12mg/100g), followed by tannin (9.61 mg/ 100g), flavonoid (6.13mg/ (4.16mg/ 100g). Alkaloids 100g), Phenol (2.34mg/100g), Cardiac glycosides (1.89 mg/100g), Phlobatannin (0.32mg/100g) and Terpenoids (0.98mg/100g). **Phytochemicals** usually possess pharmacological activities that can reduce or prevent the risk of human diseases.

3.3 Mineral Composition

Table 3 presents the result obtained for mineral analysis of the leaves of *S. anthelmia.*

| Phytochemical test | Observation | Inference |
|---|----------------------------|-----------|
| Saponins (Frothing test) | Persistence foaming | + |
| Tannins (Ferric chloride) | Greenish-brown precipitate | + |
| Flavonoids (Ethyl-acetate test) | Intense yellow coloration | + |
| Terpenoids (Chloroform and acetic anhydride test) | A grey coloration | + |
| Phenols (Distilled water test) | Blue-Greenish coloration | + |
| Alkaloid | | |
| Dragendorff's reagent | Red precipitate | + |
| Mayer's reagent | Creamy-white precipitate | + |
| Wagners reagent | Reddish-brown precipitate | + |
| Cardiac Glycoside | Brick-red precipitate | + |
| (Sulphuric acid test) | | |
| Phlobatannin | Red precipitate | + |

 Table 1. Phytochemical screening of the leaves of Spigelia anthelmia

Source: Original + Present, - Absent

Higher amounts were obtained for the macro elements, while lower amounts were indicated for micro-elements. The highest concentrations for macro-elements were observed for Calcium (2.48%), Magnesium (0.41%) and potassium (0.29%). While the highest concentration reported for micro-elements was Iron (Fe).

 Table 2. Phytochemical analysis of the leaves of S. anthelmia

| Phytochemical | Concentration (mg/100g) |
|--------------------|-------------------------|
| Alkaloid | 2.34±0.21 |
| Flavonoid | 6.13±1.08 |
| Saponin | 18.12±1.06 |
| Tannin | 9.61±0.45 |
| Phenol | 4.61±0.01 |
| Cardiac glycosides | 1.89±0.23 |
| Terpenoids | 0.98±0.51 |
| Phlobatannin | 0.32±0.27 |

Source: Original Mean ±SD, n=3

Table 3. Mineral analysis of S. anthelmialeaves

| Minerals | Concentration (%) |
|----------------|-------------------|
| Sodium (Na) | 0.022±0.03 |
| Potassium (K) | 0.23±0.12 |
| Calcium (Ca) | 0.48±0.01 |
| Magnesium (Mg) | 0.15±0.02 |
| Phosphorus (P) | 0.035±0.23 |
| Iron (Fe) | 0.19±0.02 |
| Manganese (Mn) | 0.003±0.02 |
| Copper (Cu) | 0.001±0.01 |
| Zinc (Zn) | 0.004±0.01 |

Source: Original Mean ±SD, n=3

4. DISCUSSION

Naturally occurring plant chemicals accumulate in different plant parts, such as leaves, roots, stem, bark, fruits, and seeds [11-12]. However, these plant chemicals, apart from the fact that they help in the prevention of diseases in human, they also protect the plant [13]. Apparently, all the phytochemicals examined in these studies, gave a positive result. Fagbohun et al. [14] in their findings reveal a similar result for all the phytochemical compound examined also in their study. The presence of all the phytochemicals was similarly reported by Awotedu et al. [15] in their findings on Azadirachta indica, Leea quineensis and Parkia biglobosa leaves. Alkaloids have been observed to have a microbial effect, anti-diarrheal effect, antihypertensive, antifungal, anti-inflammatory and anti-fibrogenic effect [16] Alkaloid presence was

confirmed in this study, however, the result obtained compares favourably with that reported by Awoyinka et al. [17] for alkaloid in Cnidoscolus aconitifolius. The presence of tannin was observed also in this study, meanwhile, tannin in medicinal plants shows the ability of these plants to play an essential role as antifungal, antidiarrheal, antioxidant and antihemorrhoidal agent [18]. In similar research, saponin. cardiac glycosides, flavonoids, phenolics are all present too in the seeds of Piliostigma thonningii as described by Jimoh and Oladiji [19]. A contrary result was obtained for Garg and Garg [20] where all the phytochemicals examined for Tinospora cordifolia leaves using methanolic extract gave a negative result alkaloids, cardiac glycosides. including; phenolics, and saponin except flavonoid that was present. However, the result obtained in the findings of Anyasor et al. [21] reveals the absence of some phyto chemicals like alkaloids, saponin and tannin for Costus afer. The result for the quantitative phytochemical analysis revealed some amount of all the phytochemicals present in the leaves of S. anthelmia. The saponin content of the leaves presented the highest amount of 18.12mg/100g, followed by tannin (9.61mg/100g), while the lowest value was reported for terpenoids (0.98mg/100g). Infections from parasites are usually prevented by saponins when consumed by humans. The saponin content in this study it was found to be higher compared to the values reported for Azadirachta indica, Parkia biglobosa and Leea guineensis whose values gave 15.35mg/100g, 10.47 mg/100g, 7.53mg/100g respectively as reported by Awotedu et al. [15]. The presence of saponin usually attract foamy characteristics and possess wound healing properties [22]. The values reported in these studies are all on the lower side compared to that reported by Ndukwe et al. [23] and Harry-Asobara and Samson [24]. The leaves of S. anthelmia contains a low content of alkaloids (2.34mg/100g) compared to some values reported by different authors. The values reported in the study conducted by Okeke and Ezeabara [25] for Cleome ciliata leaves, stems and roots are lower for all the phytochemicals examined compared to that reported in these studies. Terpenoids and flavonoids are present in considerable amount while alkaloids are present in very high concentration in Psidium quajava Terpenoids possess [26]. various pharmacological activities like anti-malarial, antiinflammatory, anti-cancer, anti-viral, anti-bacterial and also inhibits cholesterol synthesis [27]. The presence of phenols in the study suggests that S. anthelmia as an important plant which is responsible for the protection of tissue membranes and proteins against harmful free radicals [28-29]. Phenolic compounds are one among plant secondary metabolites that are known to act as natural antioxidants [30]. Flavonoids tend to be most commonly known with regards to antioxidant nature. They also possess anticancer, anti-inflammatory, antibacterial and anti-allergic characters [31]. They serve as potent antioxidants which can protect the human body from free radicals and reactive oxygen species. Different authors (Oluwasina et al. [32] and Dewick [33] have reported high alkaloid content in the roots other than leaves, and this is in accordance with the report of Benbott et al. [34] that high alkaloids constituent of the seed and root may arise because those organs serve as storage point of the plant. Alkaloids, apart from the fact that they are naturally derived from plants, are also produced by a large variety of organisms including bacteria, fungi, plants and animals. Alkaloids possess diverse pharmacological properties asephetrine (for antiasthma), guinine (for anti-malarial), vincamine (for vasodilatoral [35]. Alkaloids are usually known for evoking bitter taste in plants and they usually act on various metabolic systems in humans and animals [36]. Alkaloids tend to be poisonous when taken in bulk amount due to their stimulatory effects producina excitation associated with cell and nerve disorders [37]. Tannin content in the leaves of S. anthelmia was reported to be 9.61mg/100g and this is comparably higher than those reported previously in other similar studies by Awotedu et al., [15]; Senguttuvan et al., [38]; Ekwueme et al., [39]; Ajuru et al., [40]. Plants containing tannins are used for healing of wounds, antidotes for poisons, varicose ulcers, haemorrhoids, frostbite and burns Hence, herbs containing tannins can be used as mouthwashes, eyewashes, snuff and even as vaginal douches and also treat rectal disorders [41-42]. The results for the mineral composition of S. anthelmia leaves (Table 3) revealed that the leaves contain minerals in different degrees as evidenced by the values obtained. The result showed that calcium had the highest value of 0.48%, while sodium gave the lowest value (0.022%) for macro elements. In microelements. iron had the highest concentrations and all other micro elements indicate a low value. The calcium values (0.48%) obtained in this study is higher than that reported by Olaniyi et al., [43] for Crescentia cujete leaves (6.13mg/100g). Calcium is vital in the formation

of bones and teeth as well as regulation of nerve and muscle functions and blood functions [44]. Hassan et al. [45] also report that calcium is important and very safe for cancer patients. The phosphorus content (0.035%) is lower compared to the value (0.99%) reported in the findings of Awotedu et al. [15] in the leaves of Azadirachta indica which had the highest levels among the three medicinal plant examined. Phosphorus helps in the normal kidney functioning, transfer of nerve impulse and ionic balance in the body Ahmed and Chandhary [46]. However, the range (1.33-1.62) reported by Chavan et al. [47] for Phosphorus in the leaves of Artemisia nilagirica, Cythocline purpurea and Sphaeranthus indicus are very high compared to that reported in the study. A higher concentration of magnesium was observed for S. anthelmia (0.15%) compared to the value reported by Fagbohun et al. [14] for the three species examined. Magnesium is an important enzyme activator in carbohydrates and protein metabolism as claimed by Vormann [48]. The values reported for sodium and potassium are (0.022% and 0.23% respectively). The ratio of sodium to potassium should be less than 1 according to the recommendation of FND [49]. The ratio of Na: K calculated for S. anthelmia gives (0.13) which suggest that the plant would reduce high blood pressure. Ujowundu et al. [50] also reported very low values for sodium and potassium in the seeds of Dacryodes edulis. However, all the values reported the micro elements in this study are very low and are comparably similar to other results obtained by different authors.

5. CONCLUSION

The medicinal attributes of S. anthelmia leaves as evident from its ethnobotanical and folkloric uses, could be attributed to the presence of these secondary metabolites; Saponin, Tannin, flavonois, alkaloids, phenols, cardiac glycosides, terpenoids, phlobatannin. Thus, from our findings, we can suggest that S. anthelmia is a rich source of phytochemicals which in no small measure can help in the treatment, prevention and management of diverse medical complications. Also, the phytomineral status of the plant shows a considerable amount of minerals that can be beneficial to human health.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

- 1. Ajiwe VIE, Dimonyejiaku N, Ajiwe AC, Chinweuba AJ, Chendo NM. Preliminary study on the pharmaceutical constituents of *Emilia sonchifolia* leaf. Anachem Journal. 2008;2:302-309.
- Ghani A. Introduction to pharmacognosy. Ahmadu Bello University Press, Nigeria. 1990;1(2):199-205.
- Harborne JB. Phytochemical methods: A guide to modern techniques of plant analysis, 13th Ed. Chapman and Hall, Ltd. London. 1973;5-15.
- 4. Arts CW, Hollman PC. Polyphenols and disease risk in epidemiologic studies. Am J ClinNutr. 2005;81:317-325.
- Martinko MMT, John M, Parker J, Brock biology of microorganisms (8th edn.). Prentice Hall, USA; 1996.
- Olorunfemi CJ, Joseph OA, Ambrose OA, Rowland ISA. Anthelmintic efficacy of extracts of spigelia anthelmintic of extracts of Spigelia anthelmia Linn on experimental Nippostrongylus braziliensis in rats, J. Vet. Sci. 2006;7:229-232.
- Okwu DE. Evaluation of the chemical composition of indigenous spices and flavouring agents. Global J. Appl. Sci. 2001;7:455-459.
- Uzama Danlami, Osuagwu Eberechi Cecilia, Osuagwu Magnus Ifeanyi. Evaluation of the phytochemicals and antimicrobial activities of the ethanolic, Hexane and ethyl acetate extracts of *Spigelia anthelmia* leaves. International Journal of Pharmacy and Chemistry. 2017; 3(3):29-32.
- Mbaebie BO, Edeoga HO, Afolayan AJ. Phytochemical analysis and antioxidants activities of aqueous of stem bark extract of *Schotia latifolia* Jacq. Asian Pacific Journal of Tropical Biomedicine. 2012;2: 118-124.
- Association of Official Analytical Chemists (AOAC). Official methods of analysis. Food composition, additives and natural contaminants. Aldric, RC 15th edn. Association of Official Analytical Chemists Inc. USA; 2005.
- 11. Costa MA, Zia ZQ, Davin LB, Lewis NG. Chapter four: Toward engineering the Metabolic pathways of cancer-preventing lignans in cereal grains and other crops. In Recent Advances in Phytochemistry, Phytochemicals in Human Health Protection, Nutrition and Plant

Defense, ed. JT Romeo, New York. 1999;67-87.

- Zulak KG, Liscombe DK, Ashihara H, et al. Alkaloids, in Crozier A, Clifford MN, Ashihara H. (eds), Plant secondary metabolites: Occurrence, structure and role in the human diet, Blackwell Publishing Limited, Oxford, UK. 2006: 102–136.
- 13. Narasinga Rao, Bioactive phytochemicals in Indian foods and their potential in health promotion and disease prevention. Asia Pacific Journal of Clinical Nutrition. 2003; 12:9-22.
- 14. Fagbohun ED, Lawal OU, Ore ME, The proximate, mineral and phytochemical analysis of the leaves of *Ocimum gratissimum*, *Melanthera scandens*, *Leea guineensis* and their medicinal value. International Journal of Applied Biology and Pharmaceutical Technology. 2012;3: 15-22.
- Awotedu OL, Ogunbamowo PO, Emmanuel IB, Lawal IO, Phytominerals and phytochemical studies of *Azadirachta indica*, *Leea guineensis* and *Parkia biglobosa* leaves. International Annals of Science. 2019;6:28-34.
- DOI: https://doi.org/10.21467/ias 16. Ghosal S. Krishima Prasad BN. Lakshmi
- V, Antiamoebic activity of *piper longum* fruits against *Entamoeba histolytica in vitro* and *in vivo*. Journal of ethnopharmacology. 1996;50:167-170.
- Awoyinka AO, Balogun IO, Ogunnowo AA, Phytochemical screening and *in vitro* bioactivity of *Cnidoscolus aconitiifolus* (Euphorbiaceae). Journal of Medicinal Plants Research. 2007;1:63-65.
- Asquith TN, Butter LG, Interaction of condensed tannins with selected proteins. Phytochemistry. 1986;25:1591-1593.
- 19. Jimoh F.O.* and Oladiji A.T. Preliminary Studies on *Piliostigma thonningii* seeds: Proximate analysis, mineral composition and phytochemical screening. African Journal of Biotechnology. 2005;4:1439-1442.
- 20. Garg Praveen, Garg Rajesh. Qualitative and quantitative analysis of leaves and stem of *Tinospora cordifolia* in different solvent extract. Journal of Drug Delivery & Therapeutics. 2018;8:259-264.
- Godswill N. Anyasor K, Olusola Ogunwenmo, Olatunji A. Oyelana, Blessing E. Akpofunure. Phytochemical constituents and antioxidant activities of

aqueous and methanol stem extracts of *Costus afer* Ker Gawl. (Costaceae). African Journal of Biotechnology. 2010;9: 4880-4884.

- 22. Rajput SS, Soni KK, Saxena RC, Pharmacology and phytochemistry of saponin isolated from *Aloe vera* for wound healing activity. Asian J Chem. 2009;21: 1029-1032.
- Ndukwe OK, Awomukwu D, Ukpabi CF, Comparative evaluation of phytochemical and mineral constituents of the leaves of some medicinal plants in Abia State Nigeria. International Journal of Academic Research in Progressive Education and Development. 2013;2:244-252.
- 24. Harry-Asobara JL, Samson EO. Comparative study of the phytochemical properties of *Jatropha curcas* and *Azadirachta indica* plant extracts. Journal of Poisonous and Medicinal Plants Research. 2014;2:020-024.
- 25. Okeke IC, Ezeabara CA, Phytochemical screening and in vitro anti microbial activity of various parts of *Cleome ciliate* Schum. & Thonn. Bioscience Horizons. 2018;11:1-7.
- Wadood A, Ghufran M, Jamal SB, Naeem M, Khan A, Ghaffar R, Asnad. Phyto chemical analysis of medicinal plants occurring in local area of Mardan. Biochem Analyt Biochem. 2013;2:144. DOI: 10.4172/2161-1009.1000144
- 27. Mahato SB, Sen S, Advances in triterpenoid research, Phytochemistry. 1997;44:1185-1236.
- Fernandez-Panchon MS, Villano D, Troncoso AM, Garcia-Parrilla MC. Antioxidant activity of phenolic compounds: From *in vitro* results to *in vivo* evidence. Crit Rev Food SciNutr. 2008;48:649-71.
- 29. Salamah N, Ahda M, Bimantara S, Hanar R. Total phenolic content and *in vitro* evaluation of antioxidant activity of ethanol extract of *Ganoderma amboinense*. Natl J Physiol Pharm Pharmacol. 2018;8:97-101.
- Jones GA, McAllister TA, Muir AD, Cheng KJ, Effects of saponin (*Onobrychis viciifolia* Scop.) condensed tannins on growth and proteolysis by four strains of ruminal bacteria. Appl. Environ. Microbiology. 1994;60:1374-1378.
- 31. Ekam VS, Ebong PE, Serum protein and enzymes levels in rats following administration of antioxidant vitamins during caffeinated and non-caffeinated paracetamol induced hepatotoxicity.

Nigeria. Journal of Physiology Science. 2007;22:65-68.

- 32. Oluwasina OO, Olagboye SA, Olaiya A, Hassan FG, Comparative study on phytochemical quantification and antimicrobial activity of *Raufolvia vomitoria* leaves, seeds and root extracts. FUTA Journal of Research in Sciences. 2017;13: 10-16.
- Dewick PM, Medicinal Natural Products A biosynthetic approach, 2nd Ed. ed. John Wiley and Sons Ltd; 2002.
- 34. Benbott OO, Yahyia A, Belaidi A, Assessment of the antibacterial activity crude alkaloids extracted from seeds and roots of the plant *Peganu mharmala*. Journal of Natural Products and Plant Resources. 2012;2:568-573.
- Russo P, Frutaci A, Del Bufalo A, Fini M, Cesario A. Multi target drugs of plants origin acting on "Alzheimer's disease" Curr Med Chem. 2013;20:168-193.
- 36. Cushnie TP, Cushnie B, Lamb AJ, Alkaloids: An overview of their bacterial, antibiotic-enhancing and antivirulence activities. Int. J. antimicro Agent. 2014;44: 377-386.
- Jisika M, Ohigashi H, Nogaka H, Tada T, Hirota M. Bitter steroid glycosides, Vernon sides A1, A2 and A3 and related B1 from the possible medicinal plant *Vernonia amygdalina* used by wild Chimpanzees. Tetrahedron. 1992;48:625- 630.
- 38. Jamuna Senguttuvan, Subramaniam Paulsamy, Krishnamoorthy Karthika, Phytochemical analysis and evaluation of leaf and root parts of the medicinal herb, *Hypochaeris radicata* L. for *in vitro* antioxidant activities. Asian Pac J Trop Biomed. 2014;4:359-S367.
- 39. Ekwueme FN, Nwodo OFC, Joshua PE, Nkwocha C, Eluka PE, Qualitative and quantitative phytochemical screening of the aqueous leaf extract of *Senna mimosoides:* Its effect in *in vivo* leukocyte mobilization induced by inflammatory stimulus. Int. J. Curr. Microbiol. App. Sci. 2015;4:1176-1188.
- 40. Ajuru MG, Williams LF, Ajuru G, Qualitative and quantitative phytochemical screening of some plants used in ethnomedicine in the Niger Delta Region of Nigeria. Journal of Food and Nutrition Sciences. 2017;5:198-205.
- 41. Ajuru MG, Nmom FW, Oghenerukevwe OD. Qualitative and quantitative phytochemical screening of some species

of Lamiaceae in Rivers State, Nigeria. Research Journal of Food and Nutrition. 2018;2:28-37.

- 42. Norton BW. The significance of tannins in tropical animal production. Tannins in Livestock and Human Nutrition. ACIAR Proceedings No. 2000; 92:14-2.
- 43. Olaniyi MB, Lawal IO. Olaniyi AA. Proximate, phyto chemical screening and mineral analysis of *Crescentia cujete* L. leaves. Journal of Medicinal Plants for Economic Development. 2018;2: a28.

Available:https://doi.org/10.4102/ jomped.v2i1.28.

- 44. Awotedu Ogunbamowo PO. OL. Nutritional. anti-nutritional and phytochemical profile of the leaves and fruits of Synsepalum dulcificum (Schumach. & Thonn.) Daniell. American Journal of Biological Chemistry. 2019;7(3): 53-59
- Hassan LG, Abdulrahaman FN, Zuru AA, Nutritional and phytochemical investigation of *Diospyros mespiliformis* (L.)', *Nigerian* Journal of Basic Applied Science. 2004;13: 1–8.

- 46. Ahmed D, Chandhary MA. Medicinal and nutritional aspects of various trace metals determined in Ajugabracteosa. Journal of Applied Sciences Research. 2009;5:864– 869.
- Chavan YR, Thite SV, Aparadh VT, Kore BA, Quantification of minerals in some members of family Asteraceae. World journal of Pharmaceutical Research. 2014;3:613-617.
- Vormann J. Magnesium: Nutrition and metabolism. Molecular Aspects of Medicine. 2003;24:27–37. Available:https://doi.org/10.1016/S0098-2997(02)00089-4
- F. N. D. Food and Nutrition Board, Institute of Medicines. National Academy of Sciences. Dietary reference intake for energy, carbohydrate, fibre, fat, fatty acids, cholesterol, protein and amino acid (Micronutrients); 2002. Available:www.nap.edu
- Ujowundu CO, Kalu FN, Okafor OE, Agha NC, Alisi CS, Nwaoguikpe RN, Evaluation of the chemical composition of *Dacryodes edulis* (G. Don) seeds. Int. J. Biol. Chem. Sci. 2010;4: 1225-1233.

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